

## Designing a Biacore Experiment: Common Questions and Answers

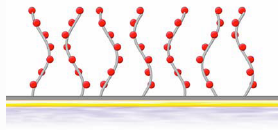
### Biacore T100 Applications: What type of experiment do you want to do?

- Specificity - Yes/No binding, Ligand fishing
- Kinetics - Kinetic rate analysis,  $k_a$  and  $k_d$
- Affinity -  $K_A$  and  $K_D$
- Concentration - How much active molecule present
- Multiple Interactions - More than one molecule binding
- Thermodynamics - How temperature affects binding (4-45°C)

### Which Chip to Use?

- CM5 - Carboxymethylated dextran matrix, most versatile chip
- CM4 - Lower degree of carboxymethylation than CM5 (less negatively charged) reduced nonspecific binding
- CM3 - Shorter carboxymethylated dextran matrix than CM5, works well with whole cells
- C1 - Flat carboxymethylated surface, no dextran matrix, works for attaching cells and viruses
- SA - CM3 chip with streptavidin
- HPA - Flat hydrophobic surface, for lipid monolayers interacting with membrane binding biomolecules
- L1 - Carboxymethylated dextran matrix with lipophilic substances, capture of liposomes
- NTA - Carboxymethylated dextran matrix with NTA, capture of His-tagged ligands

### Series S Sensor Chip CM5



- Carboxymethylated dextran matrix
- The most versatile chip available
- Excellent chemical stability

### Series S Sensor Chip C1



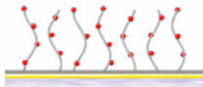
- Flat carboxymethylated surface
- For work with particles such as cells and viruses, and in applications where a dextran matrix is not desired

### Series S Sensor Chip HPA



- Flat hydrophobic surface
- For lipid monolayers interacting with membrane binding biomolecules
- Alternative to solubilization techniques for studying membrane-associated interactions
- For studies of receptors associated with membrane-like environments interacting with analytes in aqueous buffer

### Series S Sensor Chip CM4



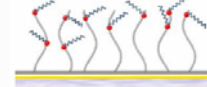
- Carboxymethylated dextran matrix with lower degree of carboxylation than CM5 (less negatively charged)
- Reduces non-specific binding of highly positively-charged molecules that may be found in cell culture, supernatants or cell homogenates
- Convenient for low  $R_{max}$  needed in kinetic applications

### Series S Sensor Chip SA



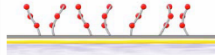
- Carboxymethylated dextran matrix pre-immobilized with streptavidin
- Captures biotinylated ligands such as carbohydrates, peptides, proteins and DNA
- Ideal for capture of biotinylated DNA fragments

### Series S Sensor Chip L1



- Carboxymethylated dextran matrix modified with lipophilic substances
- For rapid and reproducible capture of liposomes with retention of lipid bilayer structure
- No requirement for incorporation of anchoring molecules

### Series S Sensor Chip CM3



- Carboxymethylated dextran matrix
- Matrix shorter than CM5, but with the same degree of carboxylation
- For work with cells, viruses and studies of multi-component complexes
- Convenient for low immobilization levels

### Series S Sensor Chip NTA



- Carboxymethylated dextran matrix preimmobilized with NTA
- Capture of His-tagged ligands via metal chelation
- Control steric orientation of ligand component for optimal site exposure
- Generic regeneration

## What buffer should I have my samples in?

Recommended Buffers:

HBS-N - 0.1 M HEPES, 1.5 M NaCl

HBS-P - 0.1 M HEPES, 1.5 M NaCl and 0.05% v/v Surfactant P20

HBS-EP - 0.1 M HEPES, 1.5 M NaCl, 30mM EDTA and 0.05% v/v Surfactant P20

## Which molecule to immobilize (ligand)?

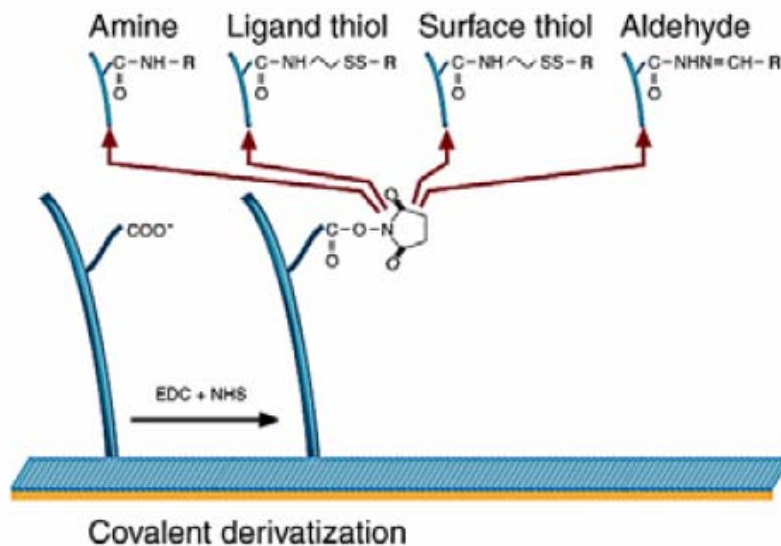
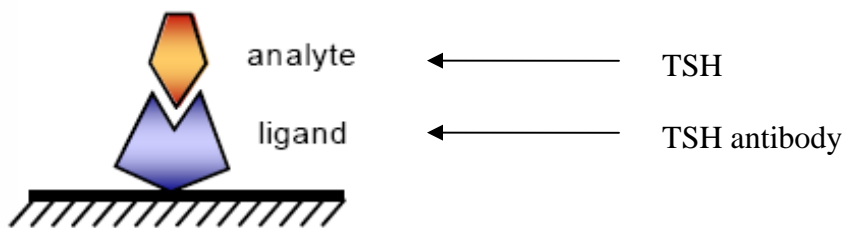
1. Immobilized molecule must maintain its functionality
2. Valency – number of binding sites (if known)
3. Charge – the molecule with the higher isoelectric point should be immobilized
4. Concentration – immobilize more “precious” sample if possible, 5-50ug/ml

5. Purity – at least 95% pure for direct coupling method
6. Tagged – tagged molecules can bind to specific chips. Ex. biotinylated ligand binding to a streptavidin (SA) chip.
7. Size of molecule - 100Da minimum to immobilize; if possible, immobilize the smaller molecule for larger signal intensities when binding the larger analyte.

### Which immobilization method to use?

Two Methods: Direct Immobilization and Capture Approach

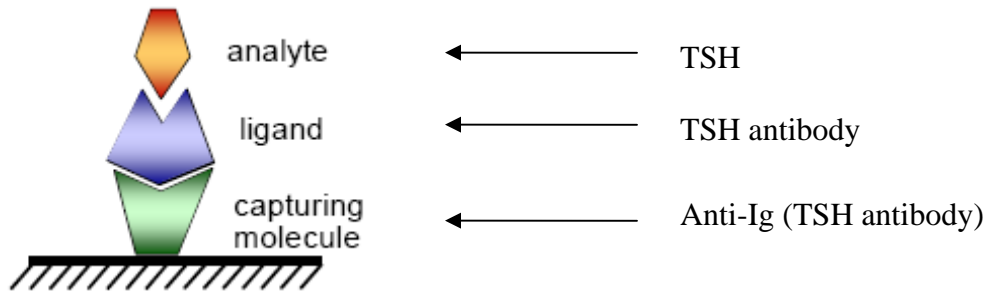
#### Direct Immobilization



Direct Immobilization chemistries:

- Amine
- Ligand thiol
- Surface thiol
- Aldehyde

## Capture Approach



Other capture molecules:

- Streptavidin : Biotin (ligand)
- anti-Biotin : Biotin (ligand)
- anti-GST : GST (ligand)
- Ni<sup>2+</sup>-NTA : 6xHis (ligand)

Advantages of capture approach:

- Can regenerate down to capture molecule so there is always a fresh surface
- Homogenous presentation of ligand
- Purification is not needed

**What do you need to know about your analyte?**

- Concentration
- Purity
- Solubility
- Appropriate buffer conditions
- Valencies